

## Table of Contents

January 2022 .....	2
Lab goals.....	2
Future To-Do .....	8

## January 2022

### Lab goals

PmrA / PriM

Re-do macrophage assay with  $\Delta$ pmrA strains

Isolate gDNA from  $\Delta$ pmrA experiment in (April?) and check by qPCR

### Saturday, January 15, 2022

#### To Do:

1. ~~Streak out chaC related cells~~
2. ~~Check glycerol stocks~~

### Results and Data:

To confirm phenotypes, streak out the following cells on CHA:

Strain Number	Description	Expected colony phenotype
-	LVS	Normal
KMLFT114	LVS $\Delta$ chaC	Mucoid
8-3	LVS $\Delta$ chaC propagated (9/5/19)	Mucoid
8-7	<del>LVS <math>\Delta</math>chaC suppressor (9/5/19)</del>	<del>Normal</del>
8-8	LVS $\Delta$ chaC suppressor (sequenced) (9/5/19)	Normal
8-9	LVS $\Delta$ chaC suppressor (9/5/19)	Normal
KRLVS42	LVS $\Delta$ chaC	Mucoid

I did NOT freeze down 8-7 and there is only a single vial of each suppressor strain. Will need to make more stocks to ship to the Mougous lab.

### Tuesday, January 18, 2022

#### To Do:

1. Image plates
2. Streak appropriate strains for stocks

### Results and Data:

Incubator wasn't on from Sat – today, so no single colonies yet. Have cold symptoms so getting COVID tested and will stay out of lab. Hannah checked on plates and the  $\Delta$ chaC\_S 8-8 plate has a bit of mold contamination, so she's re-streaking it for me.

### Thursday, January 20, 2022

#### To Do:

1. Image plates
2. Streak appropriate strains for stocks

### Results and Data:

Hannah reports that the plates only have very tiny colonies. Will image them tomorrow.

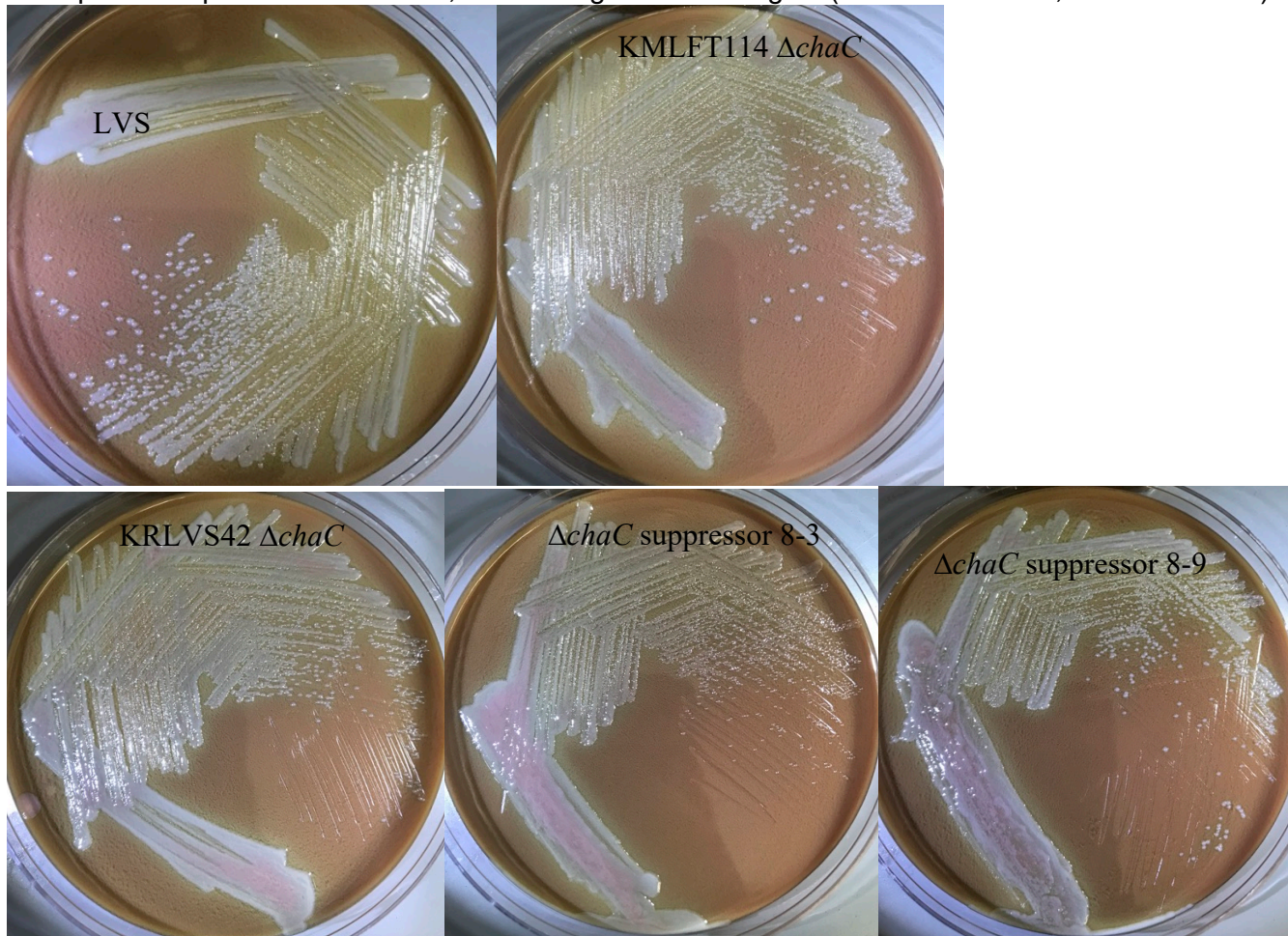
**Friday, January 21, 2022**

**To Do:**

1. Image plates
2. Streak appropriate strains for stocks

**Results and Data:**

Strains struck out over weekend have reasonable colonies, but  $\Delta chaC\_S$  8-8 still only has tiny colonies. Take photos of plates next to flame, illuminating with 2 flashlights (Hannah held one, I held the other).



Differences between LVS and  $\Delta chaC$  strains aren't huge! It looks like the  $\Delta chaC$  cells have slightly less white colonies, mucoidy isn't super evident. The differences are much clearer when comparing  $\Delta chaC\_S$  8-3 (much smaller, more transparent-looking) and  $\Delta chaC\_S$  8-9 (larger colonies, brighter white). Try imaging tomorrow to see if the differences are more evident when colonies get larger.

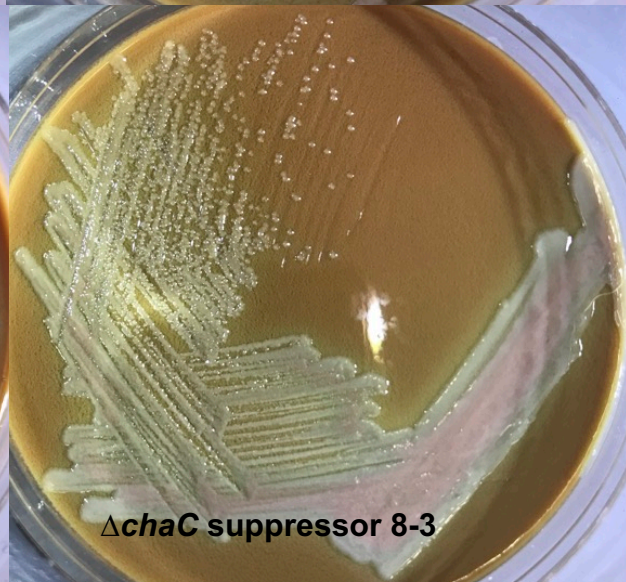
**Saturday, January 22, 2022**

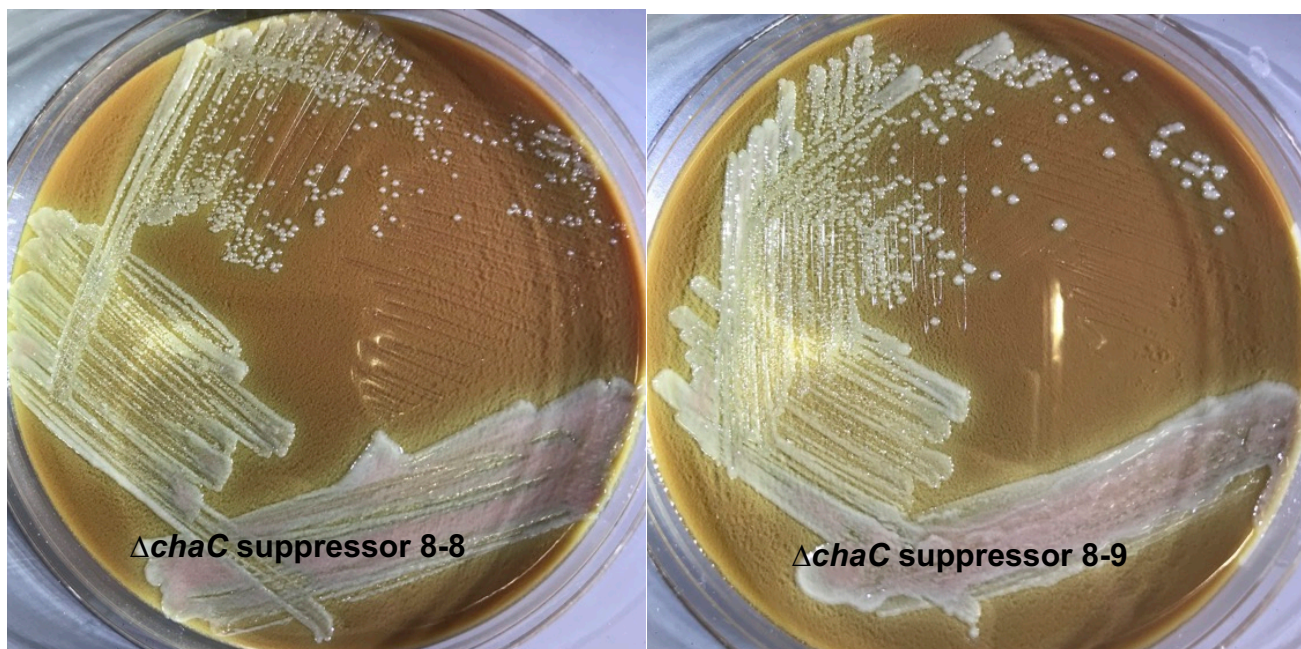
**To Do:**

1. Image plates
2. Streak out strains again

**Results and Data:**

Take more images of plates. Illuminate with brighter flashlight from yesterday and a larger powered flashlight, both propped.





Struck out strains to single colony again. Will need more CHA plates to patch out cells and make glycerol stocks to mail.

Sent email to Brook, Simon, Joseph with photos:

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Hi folks,

I've compiled some  $\Delta\text{chaC}$  information for you, below. The tl;dr version is that I can ship out the LVS, two independently generated  $\Delta\text{chaC}$  strains (slightly mucoid), and two strains with a less mucoid phenotype and one strain with a more mucoid phenotype either Tuesday or Wednesday.

#### Photographs and phenotypes:

I struck all these strains out to single colony and took photos (links attached). Note that  $\Delta\text{chaC}$  suppressor strain 8-8 had to be re-plated due to some contamination, so it's a day behind in growth. I've struck them out again to hopefully get more comparable photos. All the plates are the standard cysteine heart agar + hemoglobin plates (CHAH).

What I hope you can appreciate from the photos is that both  $\Delta\text{chaC}$  strains (KMLFT114 and KRLVS42) are a bit more mucoid than wild type LVS and have a slightly different color (less white, more green?). The mucoidy is most evident when looking at colonies next to each other- the LVS colonies stay a bit more separate while the  $\Delta\text{chaC}$  colonies look almost blurred together. The phenotype is not very obvious, but by the time I was generating the second  $\Delta\text{chaC}$  strain I had noticed it and I indicated in my notes that I could successfully predict which colonies were the deletion strain by looking at relative mucoidy.

Regarding the phenotype of the derived strains that are less or more mucoid (see isolation below),  $\Delta\text{chaC}$  suppressor 8-9 looks to me brighter white. The  $\Delta\text{chaC}$  suppressor 8-8 looks a bit in between, but that may be because it's a day behind in growth. Finally, strain  $\Delta\text{chaC}$  suppressor 8-3 appears **more** mucoid and smaller than the original  $\Delta\text{chaC}$  strains.

#### Summary of strains

LVS: non-mucoid, original strain

$\Delta\text{chaC}$  KMLFT114: original  $\Delta\text{chaC}$  strain made from LVS at CHB, slightly mucoid

$\Delta\text{chaC}$  KRLVS42: re-construction of  $\Delta\text{chaC}$  from LVS made at URI, slightly mucoid

$\Delta$ chaC suppressor 8-3: passaged  $\Delta$ chaC, more mucoid  
 $\Delta$ chaC suppressor 8-8: passaged  $\Delta$ chaC, less/non-mucoid, SNP identified (see below)  
 $\Delta$ chaC suppressor 8-9: passaged  $\Delta$ chaC, less/non-mucoid

### Isolation of the $\Delta$ chaC mucoid phenotype suppressors:

When performing the macrophage assays to test intracellular replication of the  $\Delta$ chaC cells, I was having difficulty preparing a consistent inoculum. So I performed an experiment to assess the viable CFU per OD600 for all the relevant strains in tissue culture media (DMEM+FBS). In this experiment, I included LVS and  $\Delta$ chaC cells with and without empty vector and ChaC complementation. I scraped up cells from CHAH plates, resuspended the cells in DMEM+FBS as I would for a macrophage assay, normalized OD600 to 0.03 in DMEM+FBS, 10-fold serially diluted in 1x PBS, and plated for CFU on our standard cysteine heart agar + hemoglobin plates.

In that particular experiment, I didn't find any significant differences in viable cells per CFU among the strains. But when counting the  $\Delta$ chaC colonies (no plasmid, and only these cells), I saw a mixture of normal looking colonies (opaque and white, potential suppressors) and mucoid colonies (pearly and smaller, more typical of  $\Delta$ chaC). I patched out 6 mucoid colonies and 3 normal/suppressor colonies, confirmed by PCR that all were  $\Delta$ chaC cells (no LVS contamination), and froze down 2 normal/suppressor colonies (8-8 and 8-9) and one mucoid colony (8-3).

### Whole genome resequencing

When sending out some gDNA for WGRS, I included our LVS strain, the original  $\Delta$ chaC strain (KMLFT114) and  $\Delta$ chaC suppressor 8-8. I did this sequencing at MiGS (<https://www.migscenter.com/>). As a caveat to these results, MiGS uses a Nextera kit that relies on Tn5 transposons to add adapters. When using this method with all our LVS strains, there is insertion bias that leads to extreme variability in sequencing coverage across the genome. It is formally possible that there is an undetected mutation. I think the lowest coverage we had in these samples was around 15x, so I don't think it's a strong possibility. However, it is entirely possible that there are duplications that we could not possibly detect using this method.

Compared to the reference genome, our LVS has very few mutations, nothing that appears significant.

Compared to LVS, the original  $\Delta$ chaC strain (KMLFT114) has no chaC and one slight difference from LVS: our LVS cells appear to have a mixed population of the reference sequence and a SNP in the pepN coding sequence which alters methionine 819 to isoleucine. This SNP has become fixed in the  $\Delta$ chaC cells; apart from the deletion of chaC, this is the only difference detected between LVS and  $\Delta$ chaC.

The only difference detected between the  $\Delta$ chaC cells and the chaC suppressor 8-8 is in the capA gene (FTL\_1414), a single deletion of a T in a run of Ts at bp 1343201. This is a missense mutation that leads the mutant to incorporate 5 incorrect amino acids before a stop codon. CapA is normally 403 aa, and the mutant version is truncated after 373 aa. Specifically:

LVS and  $\Delta$ chaC: 368 – Lys-Lys-Tyr-Asn-Thr-Ile...Lys-STOP (403 aa)

$\Delta$ chaC suppressor 8-8: 368 – Lys-Asn-Thr-Ile-Leu-Tyr-STOP (after 373 aa), missing 30 aa

### Confirmation by Sanger sequencing

I subsequently used Sanger sequencing to sequence ~300 bp around the SNP in LVS, the original  $\Delta$ chaC (KMLFT114),  $\Delta$ chaC suppressor 8-3,  $\Delta$ chaC suppressor 8-8, and  $\Delta$ chaC suppressor 8-9. Only  $\Delta$ chaC suppressor 8-8 had the single bp deletion in capA.

Let me know if I can provide any other information. I hope this is all helpful and I'm looking forward to hearing about what you find!

All the very best,  
Kathryn

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**Monday, January 24, 2022****To Do:**

1. Image plates
2. Streak out strains for glycerol stocks

**Results and Data:**

Words here.

**Wednesday, January 26, 2022****To Do:**

1. Make glycerol stocks
2. Ship strains

**Results and Data:**

Note included in shipping:

Strains enclosed

1 vial *F. tularensis* subsp. *holarctica* LVS

Derived from *F. tularensis* subsp. *holarctica* LVS:

1 vial KMLFT114 LVS  $\Delta$ *chaC* ( $\Delta$ FTL\_1548)

1 vial KRLVS42 LVS  $\Delta$ *chaC* ( $\Delta$ FTL\_1548)

Derived from KMLFT114 LVS  $\Delta$ *chaC*:

1 vial LVS  $\Delta$ *chaC* suppressor 8-3

1 vial LVS  $\Delta$ *chaC* suppressor 8-8

1 vial LVS  $\Delta$ *chaC* suppressor 8-9

## Future To-Do

Move 1° LVS pKR10-1 into strain box

## Bibliography

Suh, Moo-Jin et al. "Extending ribosomal protein identifications to unsequenced bacterial strains using matrix-assisted laser desorption/ionization mass spectrometry." *Proteomics* vol. 5,18 (2005): 4818-31. doi:10.1002/pmic.200402111